Effect of Creatine Supplementation and Resistance-Exercise Training on Muscle Insulin-Like Growth Factor in Young Adults

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Intr		

This study examines if creatine supplementation in resistance training individuals increases Insulin-like Growth Factor-1 (IGF-1) levels.

Conclusions

Resistance training paired with creatine supplementation does seem to increase IGF-1 levels over non-creatine supplementing resistance trainees.

Amendments

Study Design & Additional Information

The study employed 22 men and women in a creatine supplementation group that consumed a "loading" amount (16.8g) of creatine for 7 days, and then a maintenance amount (4.2g a day) for the remaining 7 weeks of the study. 20 participants were placed in the placebo group as a control to compare against, and these individuals consumed maltodextrin that was flavored similarly to creatine to mask what they were consuming. All participants had less than a year of resistance training experience. All participants were put on a similar resistance training program, with both groups performing the same amount of volume (work on the muscle) at the end of the 8 weeks (although, at the 7 week mark, creatine group did more work?).

There were no significant differences between the groups, and there was an even distribution of self-professing vegetarians and non-vegetarians. Nutrition was checked for 3 days by food diary, but no special nutrition instructions were offered other than to track their food and were taught how to do so.

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The purpose of this study was to compare changes in muscle insulin-like growth factor-I (IGF-I) content resulting from resistance-exercise training (RET) and creatine supplementation (CR). Male (n = 24) and female (n = 18) participants with minimal resistance-exercise-training experience (x1 year) who were participating in at least 30 min of structured physical activity (i.e., walking, jogging, cycling) 3–5 x/wk volunteered for the study, Participants were randomly assigned in blocks (gender) to supplement with creatine (CR: 0.25 g/kg lean-tissue mass for 7 days; 0.06 g/kg lean-tissue mass for 49 days; n = 22, 12 males, 10 female) or isocaloric placebo (PL: n = 20, 12 male, 8 female) and engage in a whole-body RET program for 8 w.K. Eighteen participants were classified as vegetarian (lacto-ovo or vegan; CR: 5 male, 5 female; PL: 3 male, 5 female). Muscle biopsies (vastus lateralis) were taken before and after the intervention and analyzed for IGF-I using standard immunohistochemical procedures. Stained muscle cross-sections were examined microscopically and IGF-I content quantified using image-analysis software. Results showed that RET increased intranuscular IGF-I content by 67%, with greater accumulation from CR (+78%) than PL (+54%; p = .06). There were no differences in IGF-I between vegetarians and nonvegetarians. These findings indicate that creatine supplementation during resistance-exercise training increases intramuscular IGF-I concentration in healthy men and women, independent of habitual dietary routine.

Errowards: pentide hormone, muscle biopsy, sport nutrition, vegetarians.

Keywords: peptide hormone, muscle biopsy, sport nutrition, vegetarians

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The combination of creatine supplementation and resistance training has been show to increase lean-tissue mass (Brose, Parise, & Tarnopolsky, 2003; Burke et al., 2000, 2003; Chrusch, Chilibeck, Chad, Davison, & Burke, 2001) and musclefiber size (Burke et al., 2003, Volek et al., 1999). The underlying mechanisms explaining the increase in muscle mass from creatine supplementation remain to be determined; however, potential mechanisms include an increase in high-energy phosphate concentration (total creatine [TCr], phosphocreatine [PCr], and creatine [Cr]; Burke et al., 2003) and PCr resynthesis after exercise (Greenhaff, Bodin, Soderland, & Hultman, 1994), cellular hydration status (Hultman, Soderland, Timmons, Cederblad, & Greenhaff, 1996), satellite-cell activity (Dangott, Schultz, & Mozdziak, 2000; Olsen et al., 2006; Vierck, Icenoggle, Bucci, & Dodson, 2003), and myofibrillar protein kinetics (Willoughby & Rosene, 2003; Parise, Mihic, MacLennan, Yarasheski, & Tarnopolsky, 2001). The might, creatine supplementation might enhance the metabolic adaptations from regular sistence-exercise training sessions, leading to greater production of insulin-like growth factor I (IGF=1) over time (Deldicque et al., 2005). This might help explain the increase in lean-tissue mass found in many creatine and resistance-exercise-training studies (Brose et al.; Burke et al., 2000, 2003; Chrusch et al.).

Most IGF-1 production occurs in the liver in response to changes in growth-

Most IGF-1 production occurs in the liver in response to changes in growthhormone concentrations and acts as an endocrine hormone, regulating tissue-specific
growth and differentiation (Czerwinski, Martin, & Bechtel, 1994; Hameed, Harridge, & Goldspink, 2002). The IGF-I produced in skeletal muscle through the
process of overload is an isoform of systemic IGF-I (Hameed et al.; MacGregor
& Parkhouse, 1996) and controls local tissue repair and remodeling (Goldspink,
1999). Borst et al. (2001) demonstrated that resistance-exercise training resulted
in a 20° increase in blood IGF-I anter 15 and 25 vecks of training in young men
and women and Sing.

1999) upported a. 500% increase in muscle stained
for IGF-I in older participants after 10 weeks of resistance-exercise training. In
1990 for our traports it was found that creating supplementation, independent of
exercise, augmented IGF-I mRNA in cultured myorubes (Louis, Van Beneden,
Dehoux, Thissen, & Francaux, 2004) and in human skeletal muscle (Deldicque
et al., 2005), possibly by enhancing the anabolic status of the cell involving IGF.
There have been several suggestions for the possible link between muscle IGF-I
activation and muscle overloading, including activation of the PI3K-Akt/PRSmTOR-signaling pathways (Deldicque et al.) and stretch tension on the basement
membrane (Goldspink) causing damage to sarcolemma and myofibrillar proteins
(Bamman et al., 2001).

Creatine supplementation results in an increase in intramuscular creatine concentrations (Green, Hultman, Macdonald, Sewell, & Greenhaff, 1996; Harris, Soderland, & Hultman, 1992). Large interindividual differences, however, in baseline resting creatine concentrations and responsiveness to creatine supplementation are evident (Casey & Greenhaff, 2000; Vandenberghe et al., 1997). Participants with initially low resting creatine concentrations (i.e., vegetarians) experience the greatest increase from creatine supplementation (Casey, Constantin-Teodosiu, Howell, Hultman, & Greenhaff, 1996; Greenhaff et al., 1994; Harris et al., 1992), leading to exercise improvements (Shomrat, Weinstein, & Katz, 2000). We have previously shown that creatine supplementation during 8 weeks of whole-body resistance-exercise training increased TCr, PCr, Type II fiber area of the vastus

lateralis, bench-press strength, and isokinetic knee-flexion and -extension work over placebo (Burke et al., 2003). Vegetarians who supplemented with creatine experienced a greater increase in TCr and PCr concentration and total isokinetic work performance over nonvegetarians (Burke et al., 2003), possibly because of

work performance over nonvegetarians (Burke et al., 2003), possibly because of lower initial resting creatine concentrations leading to accelerated intramuscular creatine uptake from exogenous supplementation.

The purpose of this study was to determine the effects of creatine supplementation (8 weeks) combined with heavy resistance-exercise training (>70% 1-RM) on muscle IGF-I concentration in vegetarian and nonvegetarian participants as previously described (Burke et al., 2003). Based on our previous findings of greater adaptations from creatine supplementation, we hypothesized that creatine supplementation during resistance training would increase IGF-I over placebo, and vegetarians on creatine would experience greater gains than nonvegetarians.

Methods

Participants

Participants

Male (n = 24) and female (n = 18) participants with minimal resistance-training experience (a lyear) who were participating in at least 30 min of structured physical activity (i.e., walking, jogging, cycling) 3–5 times a week volunteered for the study. Eighteen participants were classified as vegetarian (lacto-ovo or vegan). Participants were self-described as vegetarian, whether they were lacto-ovo or vegan, and had to have been vegetarian for a minimum of 3 years. Participant exclusion criteria included a history of creatine supplementation for 6 weeks before the start of the study or any disease or medical condition that would have prevented participation in resistance training. Participants were randomly assigned (double-blind) to receive creatine or placebo in stratified blocks based on gender. All participants completed a Physical Activity Readiness Questionnaire (PAR-Q), which screens for health problems that might present a risk with physical activity. Participants who indicated a health problem were required to have medical approval before participating in the study. The study was approved by the University of Saskatchewan ethics review board for research in human participants. The participants were informed of the risks and purposes of the study before their written consent was obtained. Participant characteristics are presented in Table 1.

Supplementation

Participants were randomized (double-blind) to supplement with creatine (loading phase; 0.25 g \cdot kg lean-tissue mass $^{-1} \cdot$ day $^{-1}$ for 7 days; maintenance phase:

Table 1 Characteristics of Participants Taking Either Creatine or Placebo, $M \pm SE$

Group	n, M/F	Age	Height (cm)	Weight (kg)	% Fat
Creatine	12/10	31 ± 2.6	170.3 ± 2.9	68.6 ± 4.0	20.5 ± 2.6
Placebo	12/8	37 + 6.8	170.2 ± 2.9	69.3 ± 4.3	22.0 ± 2.6

 $0.06~g\cdot kg$ lean-tissue mass''-day'' for an additional 49 days; n=22;~12 male [5 vegetarians], 10 female [5 vegetarians]) or placebo (maltodextrin; n=20;~12 male [3 vegetarians]). I0 female [5 vegetarians]) during 8 weeks of resistance-exercise training. The creatine loading was divided into four equal servings (-0.06 g·kg lean-tissue mass''-day'') consumed in the morning, in the afternoon or before the resistance-exercise-training session, and before going to bed. The creatine maintenance dose of 0.06~g/k kg was chosen because it has been shown to be effective for increasing muscle mass and strength (Chrusch et al., 2001). Participants were instructed to supplement with creatine immediately after each resistance-exercise-training session because creatine supplementation postexercise leads to significant muscle hypertrophy (Chilibeck, Stride, Farthing, & Burke, 2004). On nontraining days, participants were instructed to consume creatine (or placebo) in the morning or before going to bed. The average absolute daily doses of creatine for participants during loading and maintenance were $16.8\pm0.7~a$ and $4.2\pm0.2~g/d$ ay, respectively. Participants mixed each supplement with 300 ml of a fruit-flavored drink. The creatine and placebo supplements were identical in taste, texture, and appearance. Supplementation compliance was indirectly monitored by verbal communication and having participants return empty supplement bags when picking up additional supplements.

Muscle Biopsy, Histochemical Staining, and Image Analysis

Percutaneous needle biopsies were obtained from the distal third of the vastus lateralis muscle using a 5-mm Stille needle (Micrins, New York, NY) under local anesthetic with 1% lidocaine (Smith-Kline Beecham, Toronto, ON) and with suction applied via a 60-cc syringe. Participant muscle biopsies were performed 24 hr before the first training session. Target biopsy time after their last exercise session was 24 hr, with biopsies actually occurring 18–30 hr postexercise.

Preparation of staining started with fixation of the frozen section with 100% actone at 4°C for 10 min. The tissue was then washed in a bath with 10 mM of phosphate-buffered saline, pH 7.5, for 10 min. One hundred microliters of primary antibody (IGF-I: H-70, Santa Cruz Biotechnology, CA) was applied to each section and incubated for 30 min. The section was then washed with 10 mM of

Preparation of staining started with fixation of the frozen section with 100% acctone at 4 °C for 10 min. The tissue was then washed in a bath with 10 mM of phosphate-buffered saline, pH 7.5, for 10 min. One hundred microliters of primary antibody (IGF-I: H-70, Santa Cruz Biotechnology, CA) was applied to each section and incubated for 30 min. The section was then washed with 10 mM of phosphate-buffer saline, pH 7.5. Then, 100 µL of biotinylated secondary antibody (Rabbit ImmunoCruz Staining System, Santa Cruz Biotechnology) was applied and incubated for 10 min, then removed and washed well with 10 mM of phosphate-buffer saline, pH 7.5. One hundred microliters of HRP-streptavidin conjugate was then added and incubated for 10 min, which was followed by the addition of concentrated DAB chromogenic substrate and an incubation of 5 min. Then, 100 µL of hematoxylin was applied and left to sit for 2 min, which was followed by dehydration with alcohol and mounting. Six to eight samples were done at a time and always included pretraining and posttraining samples for each participant. After immunoperoxidase staining, sections were mounted, and the area positively stained was analyzed using Scion Image Version Beta 4.0.2 software (Scion Corp., Frederick, MD). First, each slide was viewed under 100x magnification (Olympus BX60, Tokyo, Japan). Then, three or four pictures were taken per slide (Spot Diagnostic Instruments Inc., Sterling Heights, MI) and immediately saved

as JPEG files on a Dell Dimension XPS R450 (Dell Computer Co., Austin, TX). Approximately 100-150 muscle fibers were used to determine the area positively stained for IGF-I content (Figure 2).

Exercise Program

All participants followed the same high-volume, heavy-load (>70% 1RM) All participants followed the same high-volume, heavy-load (27070 H8M) resistance-exercise-training program for 8 weeks. The program was a 4-day split routine involving whole-body musculature that was previously found to increase lean-fissue mass and strength (Burke et al., 2003; Candow, Chilibeck, Burke, Davison, & Smith-Palmer, 2001). Briefly, chest and triceps muscles were trained on Day 1 with the following exercises in order: flat bench press, incline bench press, flat dumbbell flies, incline dumbbell flies, cable triceps extensions, rope reverse before extensions and Fouch curls on Day 2 marticipants trained back and biceps. triceps extensions, and French curls. On Day 2, participants trained back and biceps muscles: chin-ups, low row, lat-pull downs, alternate dumbbell row, standing EZ-curls, preacher curls, and alternate dumbbell curls. Day 3 was for legs, shoulder, curs, preacher curs, and alternate dumboet curs. Day 3 was for legs, shoulder, and abdominal muscles and included the following exercises in order: vertical leg press, leg extension, hamstring curl, standing calf raises, seated dumbbell press, upright rows, shrugs, lateral raises, and abdominal crunches. Day 4 was a day of rest. These 4 days were considered one cycle, and the cycle was repeated continuously throughout the duration of the study. Participants performed seven cycles of 3–5 sets of 4–12 repetitions to muscle failure for each set. During Cycles 1 and 7, and the cycle of the cycles of 1 and 7. 3-5 sets of 4-12 repetitions to muscle failure for each set. During Cycles 1 and 7, participants performed three sets of 10-12 repetitions, with 1-min rests between sets. For Cycles 2 and 6, participants performed three sets of 8-10 repetitions, with 1.5-min rests between sets. During Cycles 3 and 5, participants performed four sets of 6-8 repetitions, with 2-min rests between sets. For Cycle 4, participants performed five sets of 4-6 repetitions, with 3-min rests between sets. Training logs detailing the weight used and number of sets and repetitions performed for each exercise were completed for every workout. Training volume was calculated (kg x reps) for the entire resistance-exercise-training program.

Diet

Dietary intake was recorded before and after the study to assess whether there Detaily make was recorded order and after the study to assess whether there were differences in total energy and macronutrient composition between creatine and placebo. Participants were given instruction about proper portion sizes and how to accurately record all food or beverages consumed. They used a 3-day food booklet to record what they ate for 2 weekdays and 1 weekend day. Fuel Nutrition software 2.1a (LogiForm International Inc., Saint-Foy, Quebec) was used to analyze the food records for total calories and the amount of energy from carbohydrate, feet and prefix and profit in the profit of the conditions. fat, and protein.

Statistical Analysis

A 2 (creatine vs. placebo) \times 2 (vegetarian vs. nonvegetarian) \times 2 (pre vs. post) ANOVA with repeated measures on the third factor was used to determine differences between the creatine and placebo groups and vegetarians and nonvegetarians

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over time. Tukey's post hoc tests were used to determine differences between group means. All results are expressed as $M\pm SE$. Statistical analyses were carried out using SPSS version 10.02 for Microsoft Windows. Statistical significance was set

Results

= .06; Figure 1).

ferences between groups for body weight or lean-tissue mass at baseline. Participants supplementing with creatine, however, experienced a greater increase in body mass and lean-tissue mass than those on placebo (body mass: CR 2.2 kg or 3.2%, Pl. 0.6 kg or 0.9%; lean-tissue mass: CR 2.5 kg or 6%, Pl. 1.9 kg or 2%; $\rho < .05$). Vegetarians on creatine experienced an increase of 2.4 kg in lean-tissue mass, compared with an increase of 1.9 kg for nonvegetarians on creatine ($\rho = .06$). Vegetarians supplementing with creatine experienced a greater increase in

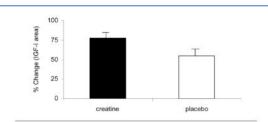


Figure 1 — Change in area positively stained for insulin-like growth factor-I (IGF-I) from before to after training and supplementation, $M\pm SE$ (p=.06). Area is expressed in μm^2

Figure 1: The researchers took muscle biopsies (cut out a tiny bit of muscle) from participants of the study and then probed (stained) them for Insulin-like Growth Factor (IGF) and compared the amount of IGF present between creatine supplementing participants and placebo (control group - no creatine given) before and after resistance training. This graph is the amount of difference between before and after.

- Although not shown in the graph, the placebo group had a 55% increase in IGF, while the creatine group had a 78% increase in IGF.

 Technically, at a p-value (significance value) of 0.05, these data are non-significant, but the p value is 0.06 which seems highly likely these are meaningful differences between the two

Take Away: Creatine supplementation likely increases IGF on muscle after resistance

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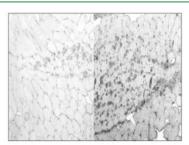


Figure 2 — An image of one participant's muscle cross-section stained for insulin-like growth factor-I presupplementation (left) and after creatine supplementation (right).

high-energy phosphate content than nonvegetarians on creatine (TCr: vegetarians 25%, nonvegetarians 7%; PCr: vegetarians 37%, nonvegetarians 11%; p < .05). There were no changes in TCr, PCr, or free Cr for placebo participants. Creatine supplementation increased Type II fiber area of the vastus lateralis by 28% (p < .05), compared with a 5% increase for placebo. The change in lean-tissue mass was significantly correlated to the change in intramuscular TCr content (r = 61, p < .05), and the change in r = 100 (r = 100).

Discussion

The primary purpose of this study was to determine the effects of creatine supplementation and resistance-exercise training on muscle [GF-I in young adults, Results showed that muscle [GF-I content was significantly increased after high-intensity resistance-exercise training, with greater gains observed from creatine supplementation than from placebo. [GF-I has been shown to increase muscle protein synthesis and satellite-cell activity (Allen & Boxhorn, 1989) and stimulate the PJ3K-Akt/ PKB-mTOR-signaling pathway involved in muscle hypertrophy (Deldicque et al., 2005). In the current study, participants supplementing with creatine experienced a greater increase in [GF-I han those on placebo (CR 78%, PL.55%; Pp. 90,) These results support the findings of Deldicque et al., who observed a 30% increase in IGF-I mRNA expression at rest after 5 days of creatine supplementation in young adults. Our results further suggest, however, that regular resistance-exercise-training sessions for 8 weeks increase muscle IGF-I in adult humans, with greater gains observed from creatine supplementation. Although the mechanism explaining the

Figure 2: The researchers took muscle biopsies (cut out a tiny bit of muscle) from participants of the study and then probed (stained) them for Insulin-like Growth Factor (IGF) and compared the amount of IGF present between creatine supplementing participants and placebo (control group - no creatine given) before and after resistance training. This image is a 'cross section' of a person's muscle - the black dots are the staining for IGF before resistance training (left) and after resistance training (right).

Nothing new here that wasn't discussed in Figure 1.

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increase in IGF-I from creatine remains to be elucidated, the most plausible theory involves high-energy phosphate metabolism and training intensity. As we have previously shown, creatine supplementation increased both PCr and TCr content to a greater extent than placebo (Burke et al., 2003). The increase in high-energy phosphate metabolism might have allowed resistance training to be performed with greater intensity as was observed in Weeks 2 and 7 of our resistance-exercise-training program. The higher metabolic demand from more-intense resistance-exercise-training sessions might explain the greater increase in muscle IGF-I content from creatine supplementation found in the current study.

It is unclear why vegetarians did not experience a greater increase in IGF-I than

It is unclear why vegetarians did not experience a greater increase in IGF-1 than nonvegetarians. It has been shown that habitual dietary intake of reduced energy and protein might reduce serum IGF-1 in humans (Thissen, Ketelslegers, & Underwood, 1994). In particular, a diet low in essential amino acids reduces IGF-1 production (Harp, Goldstein, & Phillips, 1991), suggesting that essential amino acids are necessary to maximize IGF-1 production. For the current study, vegetarians consumed approximately 2,200 kcal and 79 g of protein per day, compared with 2,650 kcal and 139 g of protein per day, compared with 2,650 kcal and 139 g of protein per day for nonvegetarians. Although we cannot differentiate between essential and nonessential amino acids that have been shown to blunt IGF-1 production (Clemmons, Seek, & Underwood, 1985) and might have contributed to our lack of significant findings.

diets tend to be low in one or more essential amino acids that have been shown to blunt IGF-1 production (Clemmons, Seek, & Underwood, 1985) and might have contributed to our lack of significant findings.

As previously reported, creatine supplementation resulted in greater increases in lean-tissue mass and Type II fiber area than placebo (Burke et al., 2003). It is difficult to determine whether the greater increase in lean-tissue mass and fiber area with creatine was caused by greater muscle protein accretion. There was a trend (p = .06) for a greater increase in muscle IGF-1 content with creatine supplementation than with placebo, suggesting a greater muscle protein synthetic response from creatine and exercise. The greater intramuscular IGF-1 content (78%) from creatine supplementation than with placebo (55%) might help explain the differences in muscle mass and exercise performance as previously reported (Burke et al., 2003). The addition of creatine and subsequent increase in TCr and PCr might have directly or indirectly stimulated production of muscle IGF-1 concentration and muscle protein synthesis, leading to muscle hypertrophy.

In summary, a structured resistance-exercise-training program increases

In summary, a structured resistance-exercise-training program increases IGF-I content in men and women. The addition of creatine further augments the physiological adaptations from resistance training, with no differences between vegetarians and nonvegetarians. Future research should determine the mechanisms explaining hormonal changes resulting from creatine supplementation alone and in combination with resistance-exercise training.

Acknowledgments

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